

Development of a system of early detection of zebra mussel through analysis of eDNA

Fast monitoring of the presence of IAS on river basins



Marta Prado Rodríguez

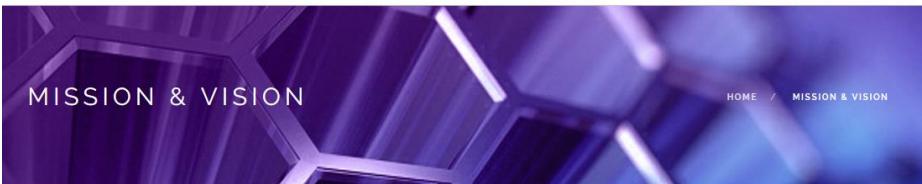
Dep. of Life Sciences/Nano4Food Unit/Food Quality and Safety Research Group

- ✓ Presentation
- ✓ What is environmental DNA and Why we use it?
- ✓ Our approach to detect eDNA
 - ✓ DNA purification module
 - ✓ DNA amplification module and protocol
 - ✓ DNA detection module
 - ✓ Data acquisition
- ✓ Final considerations

ABOUT INL

The INL International Iberian Nanotechnology Laboratory, located in Braga (North of Portugal) was founded by the governments of Portugal and Spain under an international legal framework to perform interdisciplinary research, deploy and articulate nanotechnology for the benefit of society. INL aims to become the world-wide hub for nanotechnology addressing society's grand challenges.

The INL research programme comprises four strategic fields of application of nanoscience and nanotechnology: Food and Environment monitoring, ICT, Renewable Energy and Health.

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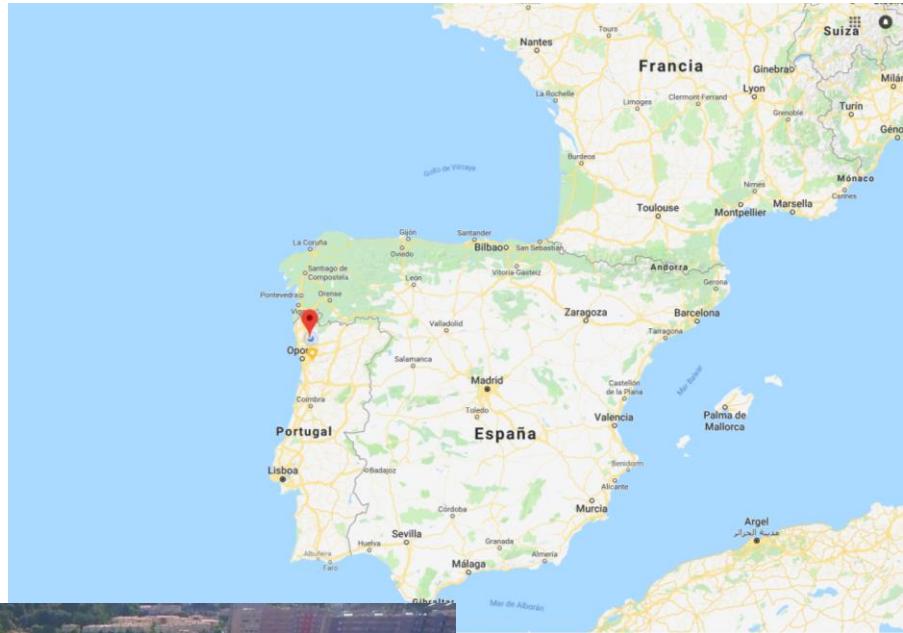
MISSION

Perform interdisciplinary cutting edge research, deploy and articulate nanotechnology for the benefit of Society.



VISION

Become the worldwide hub for nanotechnology deployment, addressing society's grand challenges.



What is environmental DNA and Why we use it?

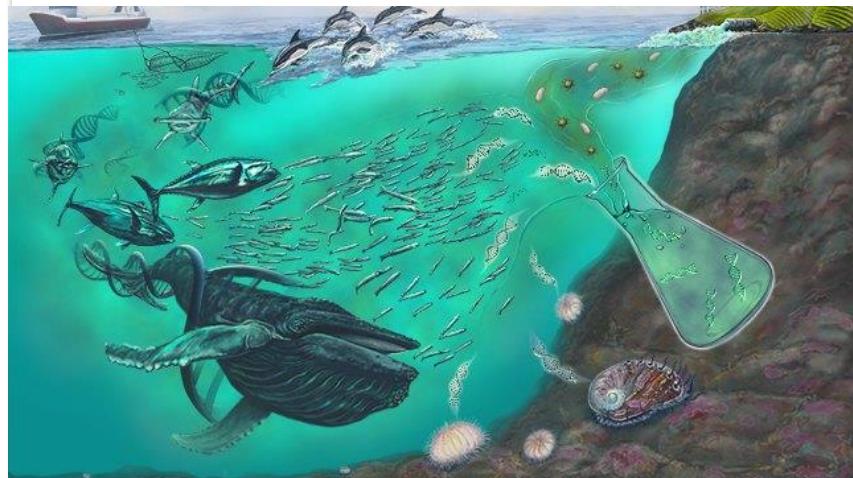
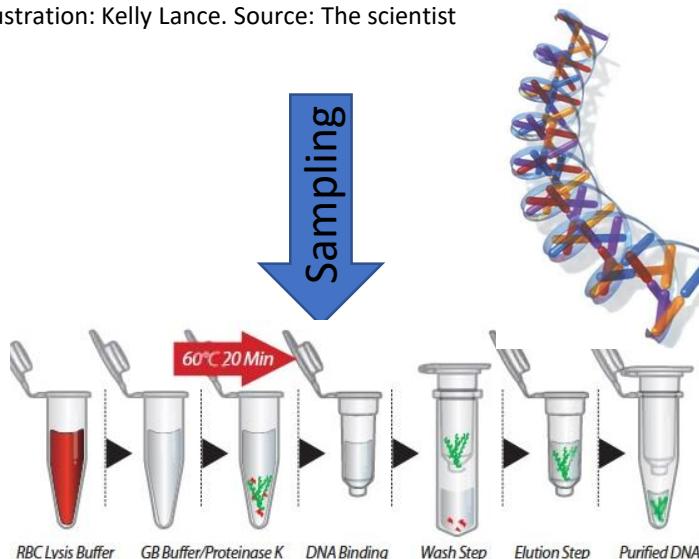
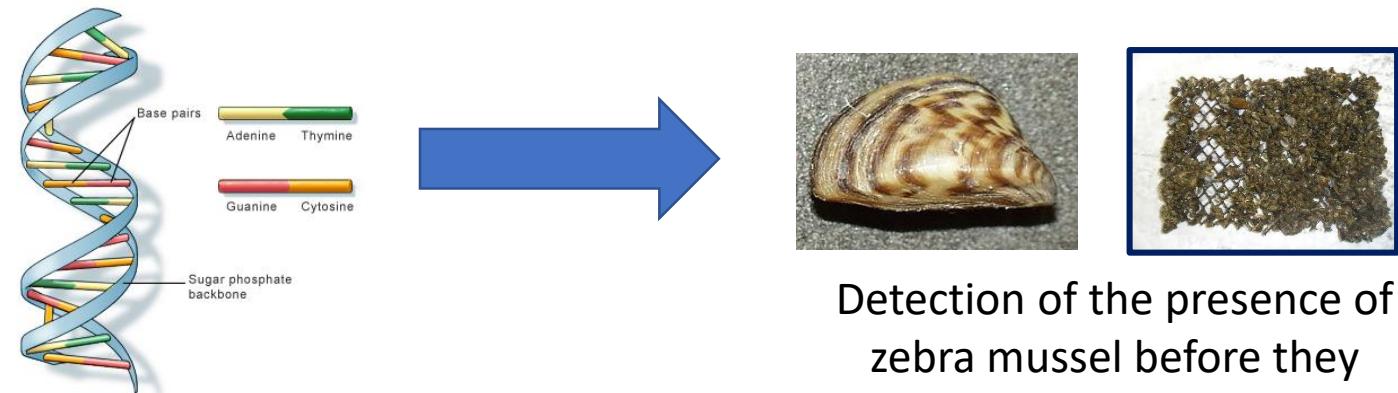


Illustration: Kelly Lance. Source: The scientist



eDNA extraction and concentration

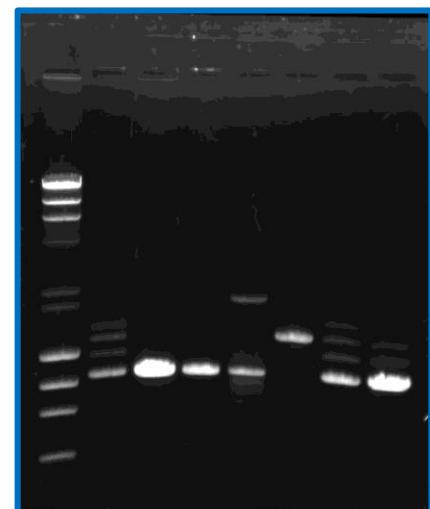


Detection of the presence of zebra mussel before they produce noticeable harm

Conventional or “bench-top” approach



Species-specific DNA amplification



DNA detection

Our approach to detect eDNA

Cooperation agreement INL-CHG for the development of system of early detection of zebra mussel through analysis of eDNA



DNA NANO.SCREEN

We use a modular design and evaluation to develop reliable analytical solutions

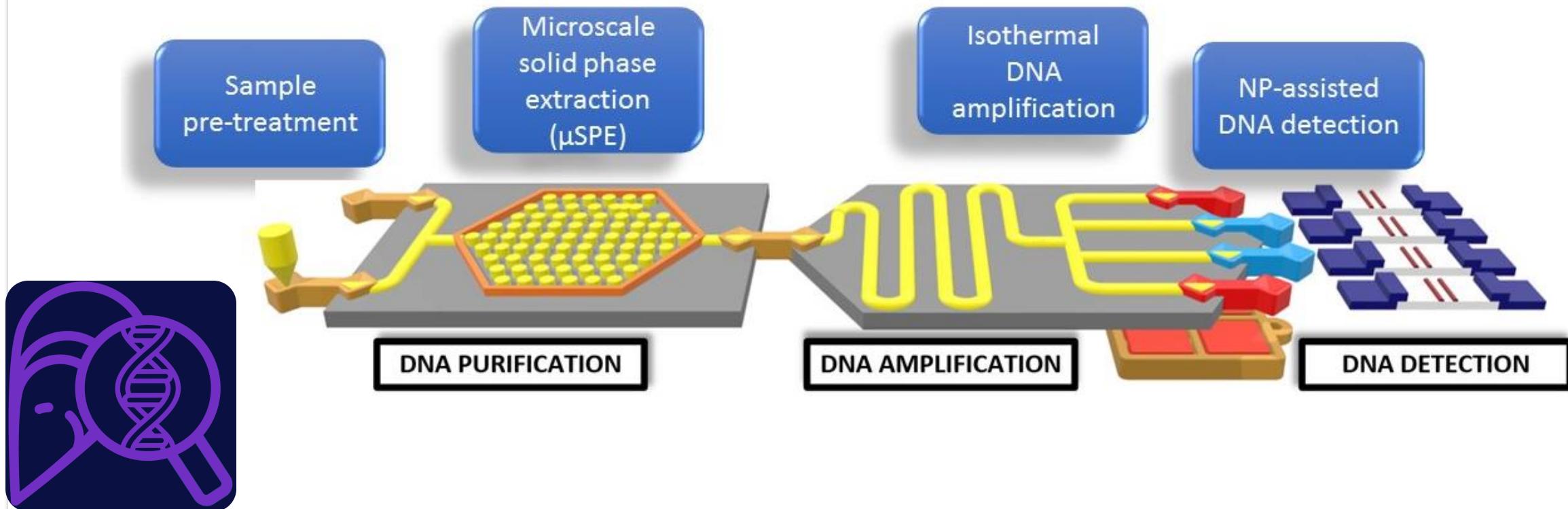
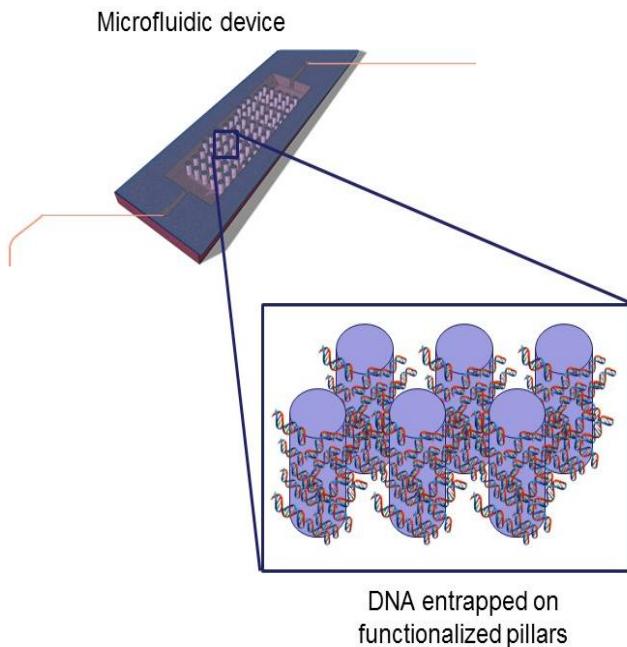
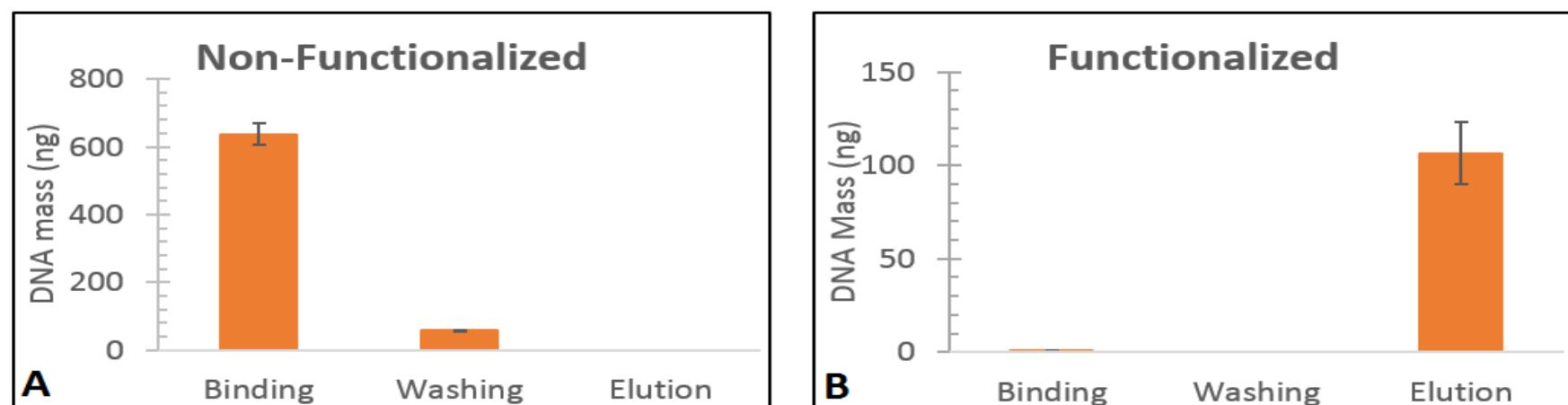




Fig 1. Microfluidic device for DNA purification

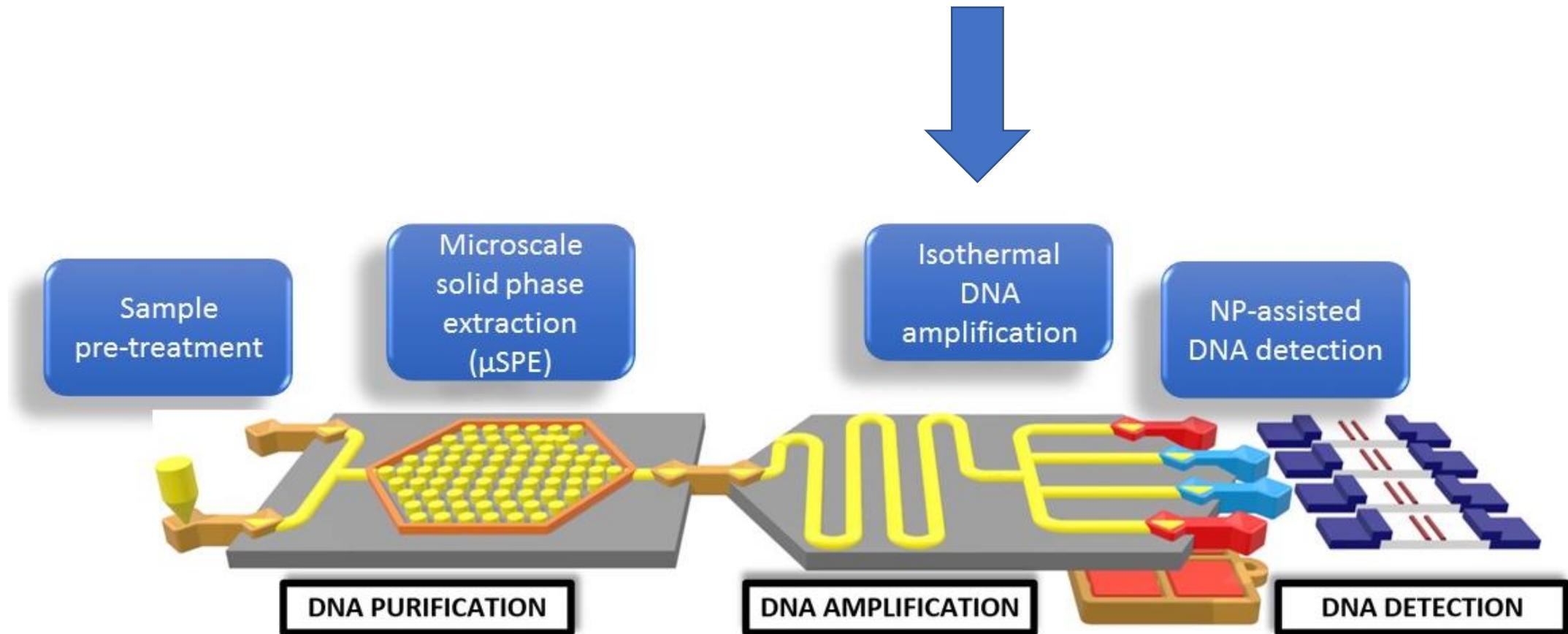


High efficient DNA extraction and purification from complex matrixes combining micro Solid Phase Extraction (μ SPE) and microfluidics



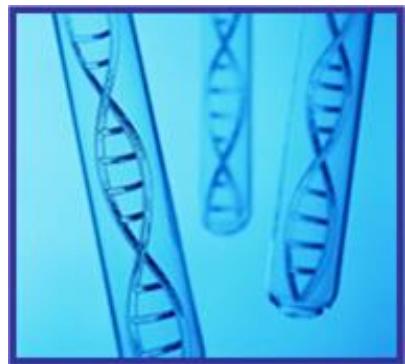
DNA purification with a non-functionalized device (A) and a functionalized device (B) using the same pH-induced DNA capture/release protocol.

Our developments

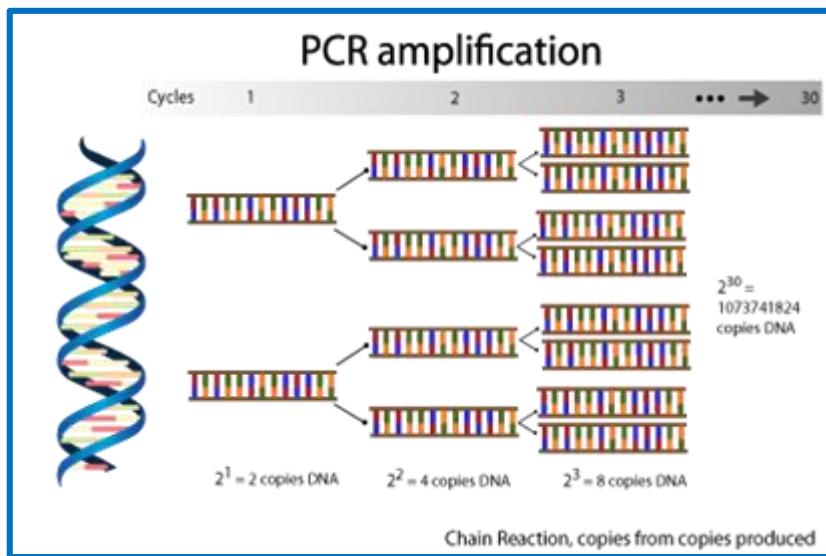


DNA amplification module and protocol

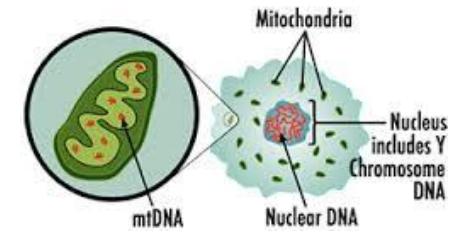
- **2 specific methods developed and tested:**
 - qPCR based approach for the specific amplification and detection of zebra mussel
 - LAMP based approach (isothermal amplification)



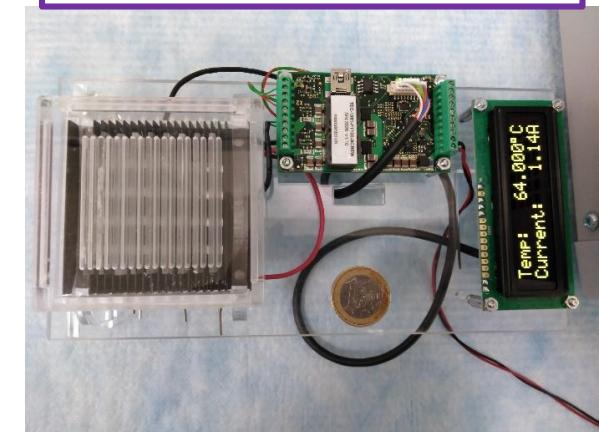
Total DNA extract



Polymerase chain reaction (PCR): exponentially amplify a single copy or a few copies of a specific segment of DNA to generate thousands to millions of copies of a particular DNA sequence



Miniaturized isothermal amplification device with integrated thermal control



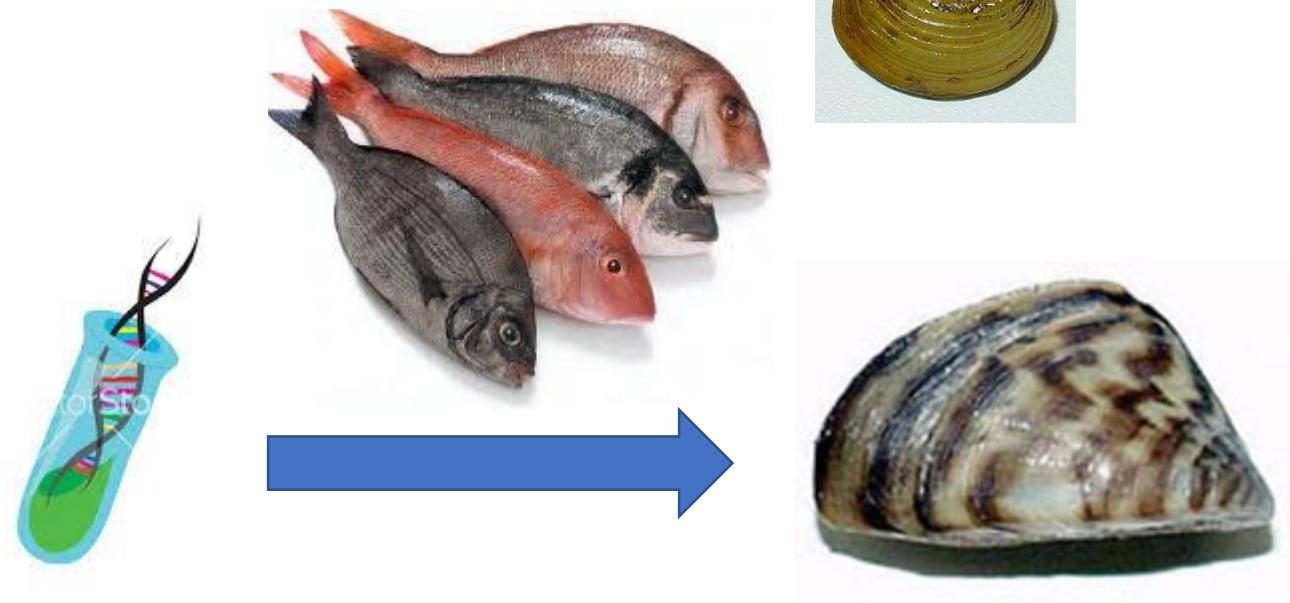
Protocols tested with:

17 well characterized autochthonous species from the Guadalquivir (Guadalictio/ Alphanius Research Group. U Córdoba) and Asian clam from Loire river (USC)



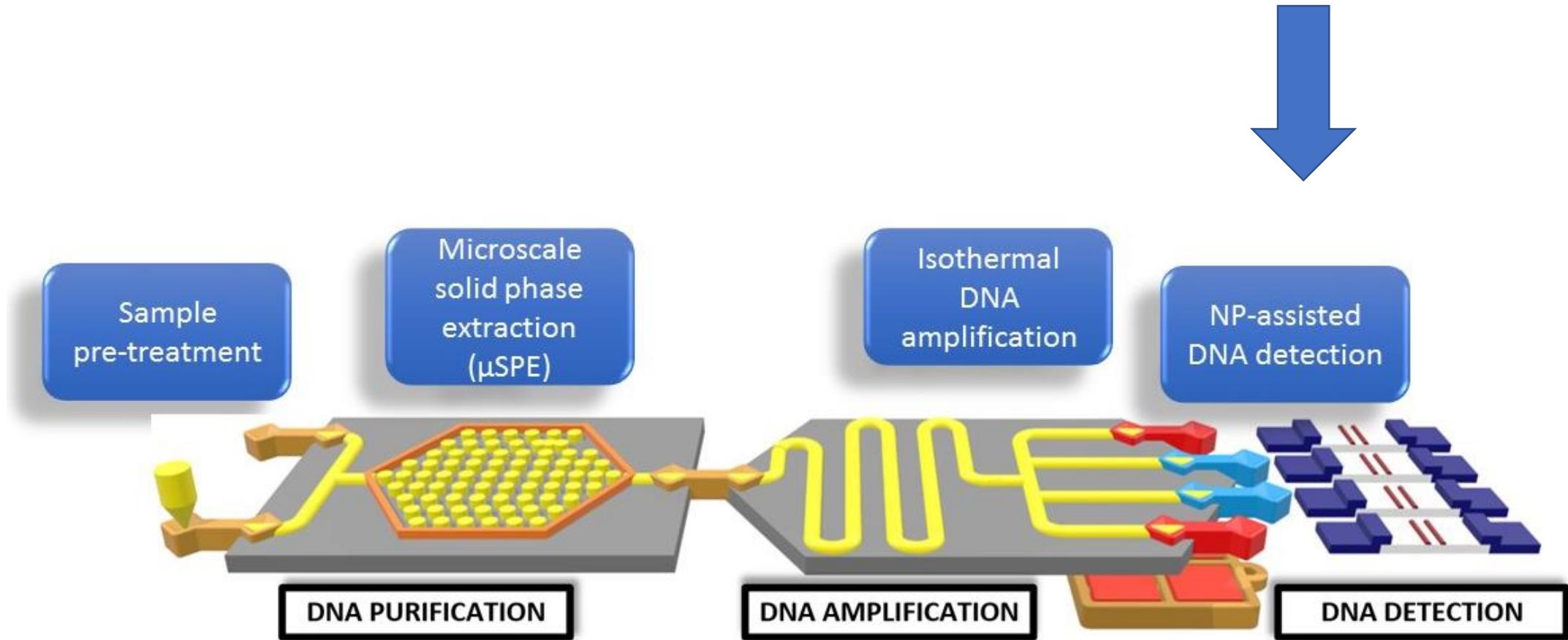
Sensitivity:

PCR up to 0.056 pg/mL
LAMP up to 0.56 pg /mL

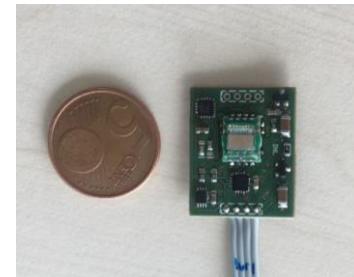
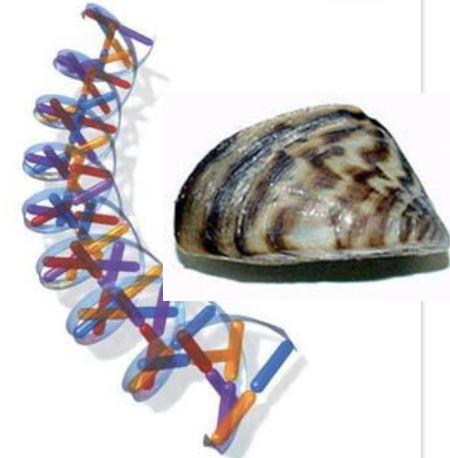
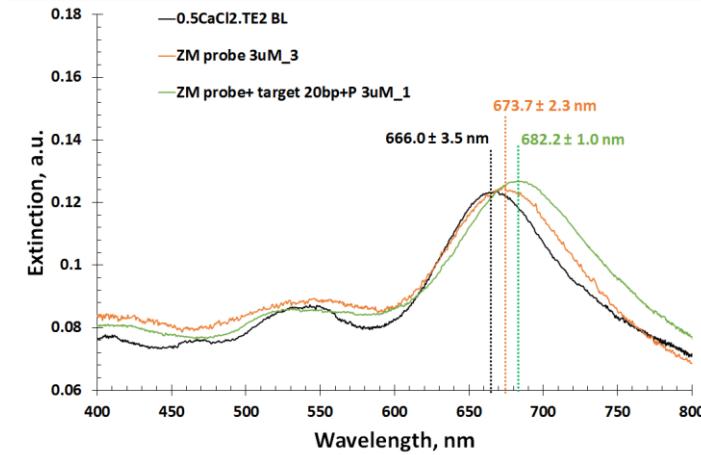
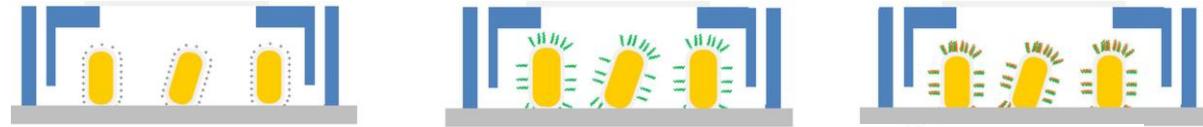
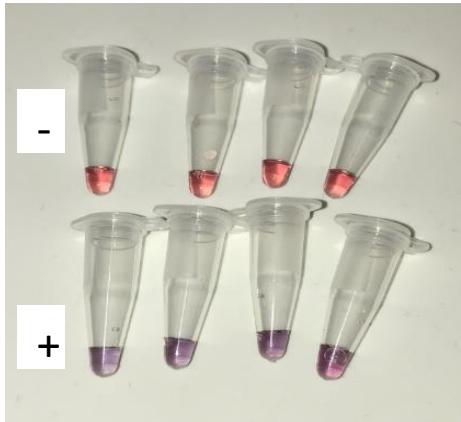
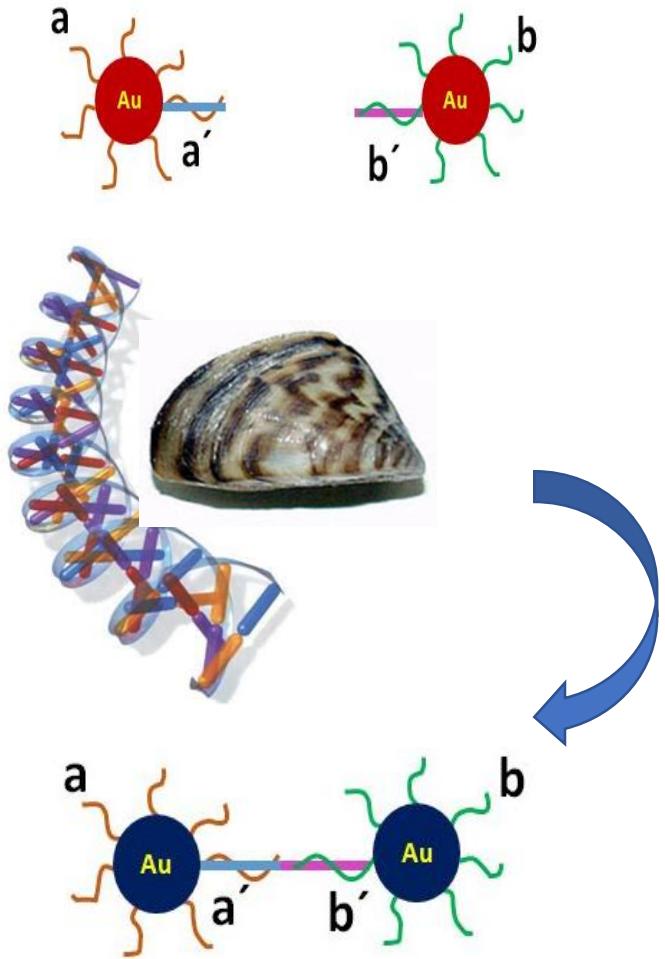


	SAMPLE	STANDARD INTERNATIONAL NAME	NR. OF SAMPLES	SAMPLING LOCATION	qPCR		qLAMP
					Hydrolysis Probe	F3/B3	
NEGATIVE SAMPLES	Fish	<i>Luciobarbus sclateri</i>	Southern Iberian barbel	3	Guadquivir river (Spain)	-	-
		<i>Pseudochondrostoma willkommii</i>	Southern straight-mouth nase	2		-	-
		<i>Squalius alburnoides</i> complex.	Calandino	3		-	-
		<i>Squalius pirenaicus</i>	Southern Iberian Chub	3		-	-
		<i>Cobitis poaludica</i>	Southern Iberian spined-loach	3		-	-
		<i>Iberochondrostoma lemmingii</i>	Iberian arched-mouth nase	3		-	-
		<i>Cyprinus carpio</i>	Carp	1		-	-
		<i>Carassius gibelio</i>	Prussian carp	3		-	-
		<i>Lepomis gibbosus</i>	Pumpkinseed	3		-	-
		<i>Micropterus salmoides</i>	Largemouth Black-bass	3		-	-
		<i>Gambusia holbrooki</i>	Eastern mosquitofish	3		-	-
		<i>Alburnus alburnus</i>	Bleak	3		-	-
		<i>Corbicula fluminea</i>	Asian clam	1	Loire river (France)	-	-
		<i>Physa acuta</i>	Freshwater snail (general)	3	Guadquivir river (Spain)	-	-
		<i>Ancylus fluviatilis</i>	Limpet (general)	3		-	-
		fam. Unionidae	Freshwater mussel (general)	4		-	-
POSITIVE SAMPLES	Crustacean	<i>Procambarus clarkii</i>	red swamp crayfish	1	Guadquivir river (Spain)	-	-
	Meat Samples	D. polymorpha 1	Zebra mussel	1	Guadquivir river (Spain)	+	+
		D. polymorpha 2		1		+	+
		D. polymorpha 3		1		+	+
		D. polymorpha 4		1		+	+
		D. polymorpha 5		1		+	+
	Positive Water Samples	D.p. transport water 1	**	1	Guadquivir river (Spain)	+	+
		D.p. transport water 2		1		+	+
		D.p. transport water 3		1		+	+
		D.p. transport water 4		1		+	+
		D.p. transport water 5		1		+	+
		D.p. transport water 6		1		+	+
		Surface river water 1 (Los Bermejales, December 2016)		1		-	-
		Surface river water 2 (Guadix, May 2017)		1		+	+

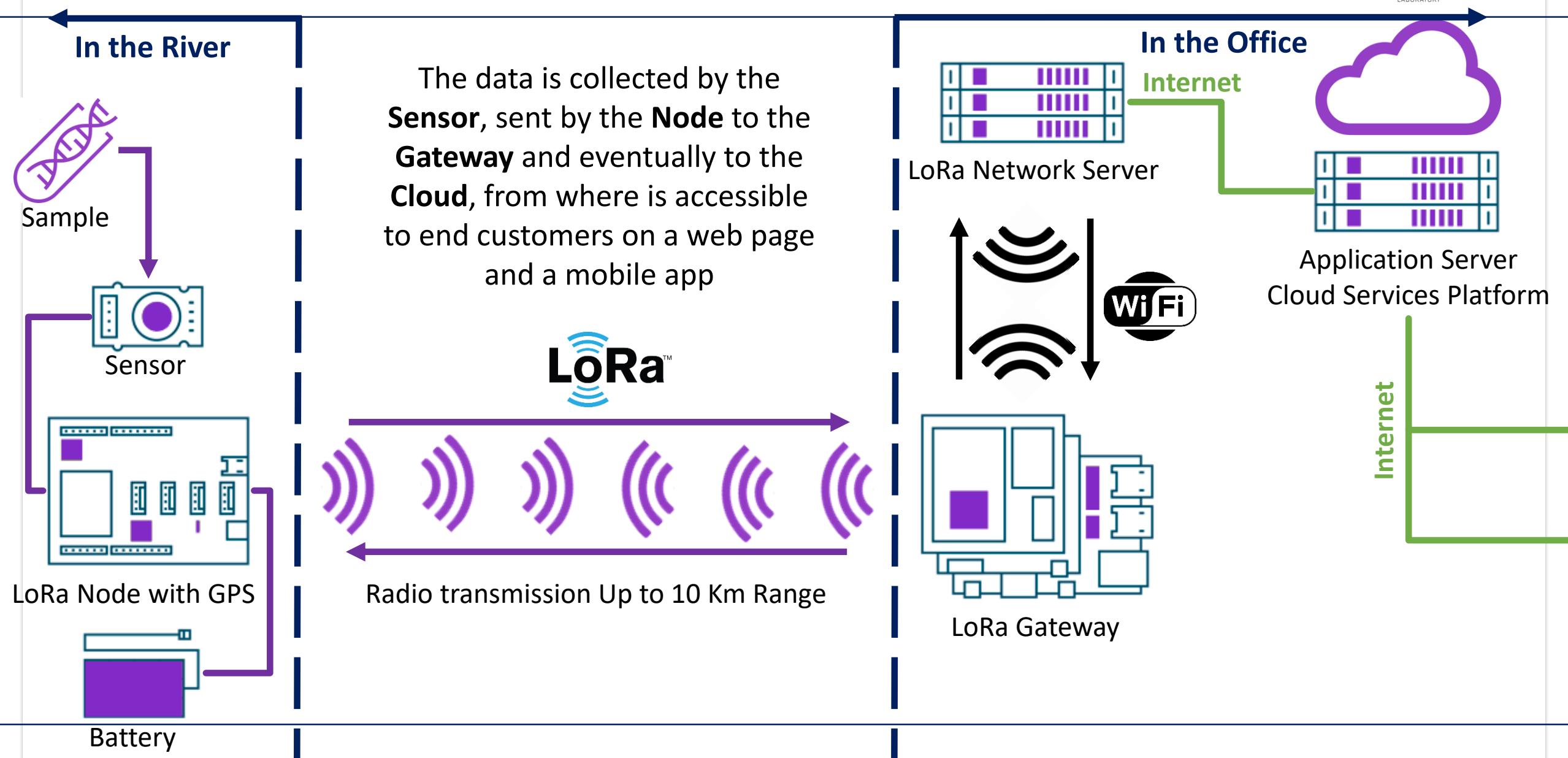
DNA detection module



DNA detection module



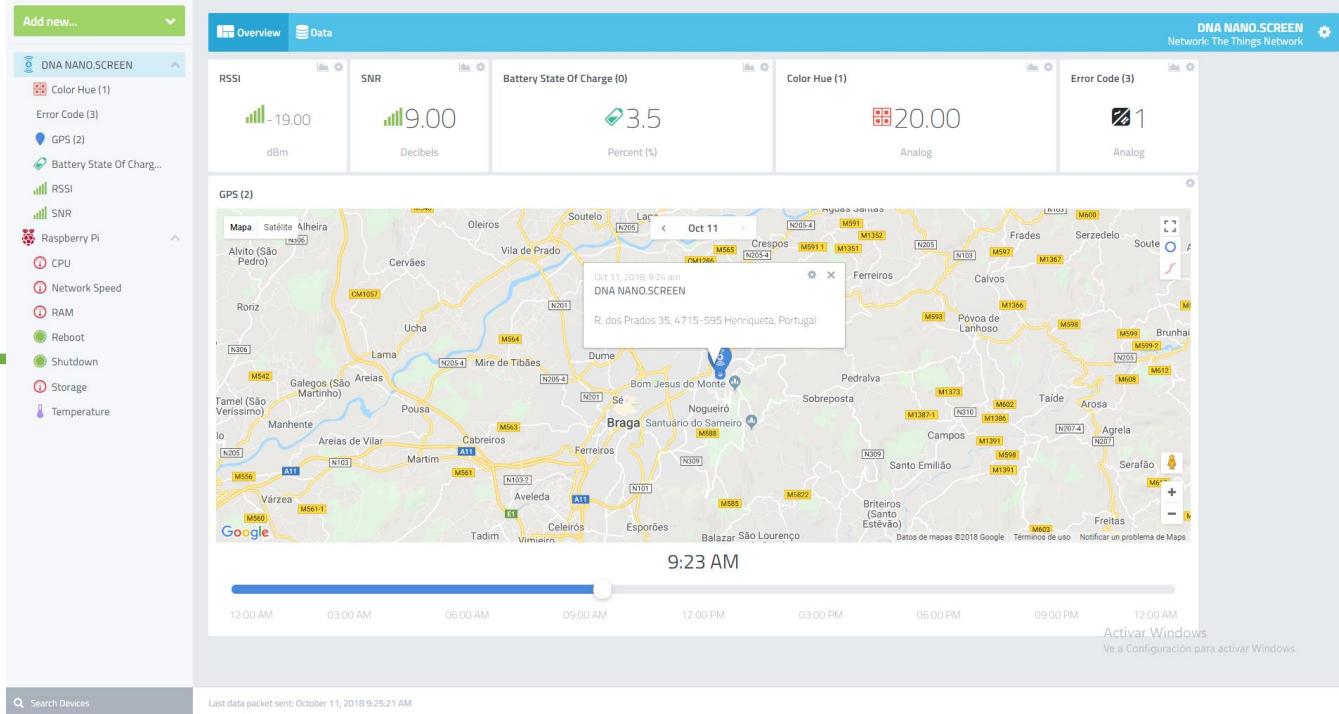
Data acquisition



Data acquisition

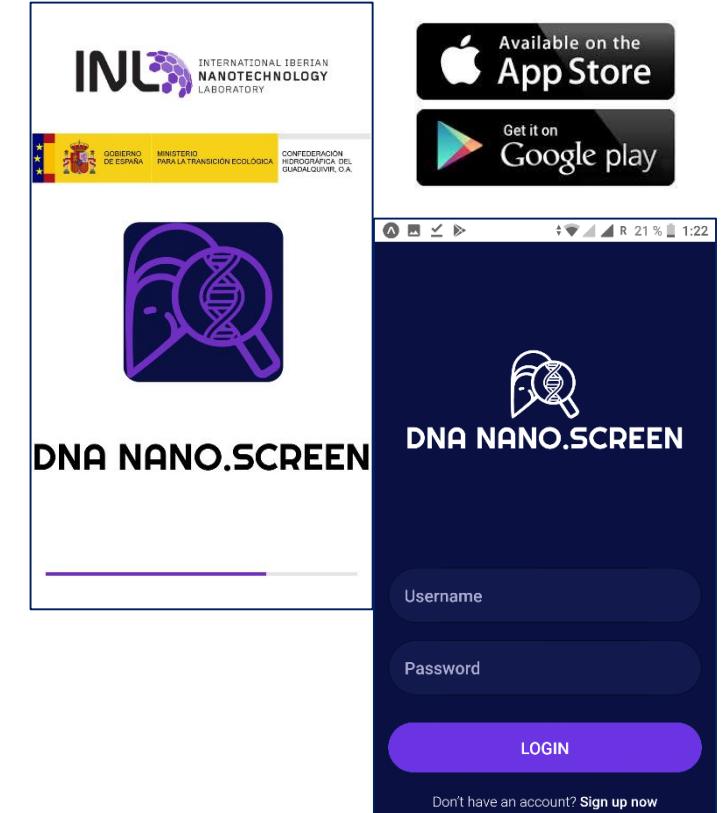
In the Office

On a web page accessing from my PC



Everywhere.....

With an app at my smartphone



- Evaluation of each individual module allow us to develop the most efficient approach in each case
- Integration of the independent modules will allow the development of automated in-situ monitoring system
- Real time sample data adquisition, transmission and accesibility together with the GPS location will allow the early identification of the possible presence of zebra mussel and geographical traceability of the results

Thanks for your attention

Joana Carvalho

Dr. Alejandro Garrido

Dr. J. Rafaela Guerreiro

Monisha Elumalai

Dr. Andrey Ipatov

Dr. Lorena Diéguez

Yosbel Toledo



DNA
NANO SCREEN

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Guadalictio/ Alphanius Research Group. (U Córdoba)

Aguas de Córdoba

Dr. Manuel Rey Méndez (USC)
Dr Jorge Barros-Velázquez (USC)

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Thank you for listening!

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